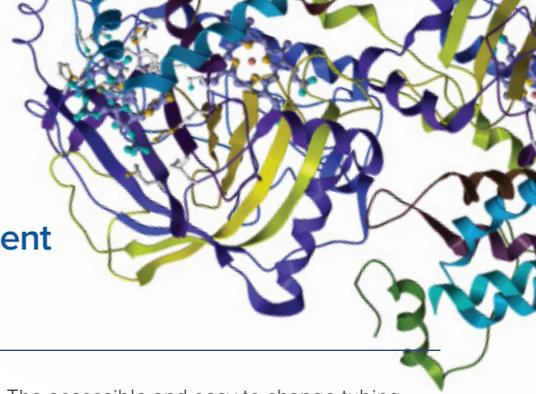


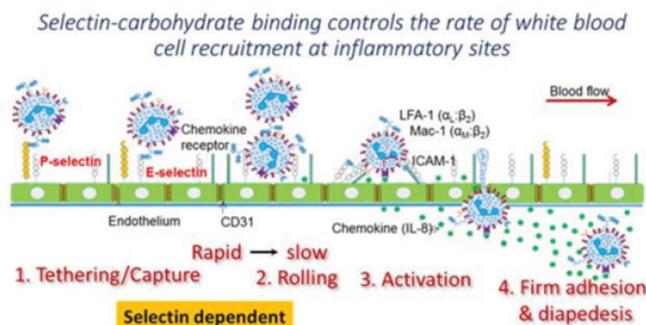
REICHERT SPR SYSTEMS

APPLICATION NOTE 17

Reichert SPR Measuring White Blood Cell (Leukocyte) Recruitment Onto Substrates Bearing Vascular Adhesion Molecules



The application of Surface Plasmon Resonance (SPR) for cell based assays is an emerging research theme. The accessible and easy to change tubing (typical i.d. values ranging from 0.005-0.04" (125-1000 μm)) and ability to change flow rate over a wide range (0.1 $\mu\text{L}/\text{min}$ to 3 mL/min) makes Reichert SPR instruments particularly well suited for cell studies. This application note demonstrates the use of the Reichert SR7500DC SPR instrument to measure the binding of white blood cells or leukocytes (analyte) to substrates bearing P-selectin (target). Commonly, such molecular binding occurs at sites of inflammation where carbohydrate ligands expressed on the leukocyte surface bind selectins expressed on the inflamed vessel wall (see figure below). This enables leukocyte recruitment at sites of inflammation, and the same process is also important during normal physiological immunity.



Experimental

BACKGROUND

Sensor Chip: Reichert Planar mixed SAM

Running Buffer: PBST

Anti-human IgG (MW 150 kDa, concentration 80 $\mu\text{g}/\text{mL}$ in pH 6.0 acetate buffer) was amine coupled to the left channel only by injecting for 10 minutes at a flow rate of 15 $\mu\text{L}/\text{min}$ after EDC/NHS activation. Baseline increased by 200-250 μRIU . Recombinant P-selectin-Fc (5 $\mu\text{g}/\text{mL}$; 10 min. capture at 15 $\mu\text{L}/\text{min}$; 240 kDa) was then immobilized on the left channel to ~ 400 μRIU . This protein contains the N-terminal lectin binding domain of P-selectin fused to a human IgG1 Fc tail. Such immobilization is consistent with 1:1 antibody:protein capture interaction. Both channels were blocked with 2% BSA (Bovine Serum Albumin). Subsequently, 107 /mL human leukocyte HL-60 cells were perfused in both channels at 50 $\mu\text{L}/\text{min}$.

Blocking studies were performed in some runs by incubating HL-60 cells with a blocking monoclonal antibody KPL-1 that binds

the N-terminal portion of the glycoprotein ligand PSGL-1 (CD162, P-selectin glycoprotein ligand-1). This is the major ligand binding epitope of P-selectin on the human leukocytes.

In other controls, the substrates contained a fusion protein called 19Fc, an irrelevant IgG1 fusion protein similar in all aspects to P-selectin-Fc except that the lectin-domain of selectins is replaced by 19 amino acids.

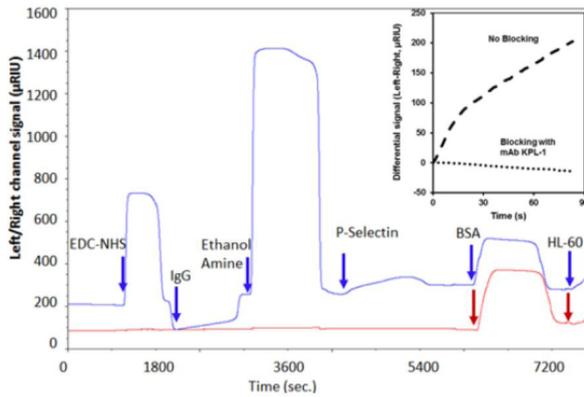


Figure 1

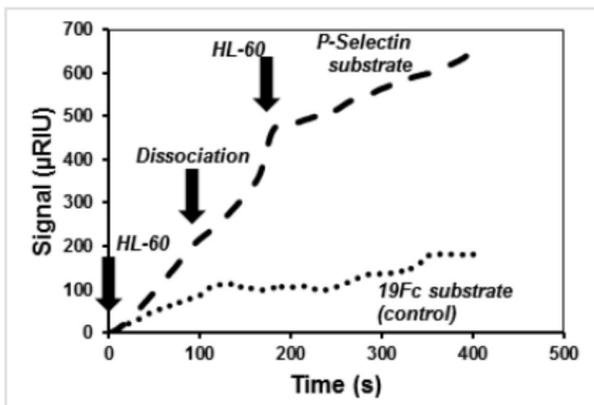


Figure 2

Results

Cell capture is noted in the left but not right channel in Fig. 1. Cell adhesion to immobilized P-selectin was prevented by the addition of the blocking antibody KPL-1.

Figure 1. Sensorgram shows the coupling of P-selectin onto the sensor substrate and subsequent capture of leukocytes on the left channel (blue). There is minimal signal change in the right channel upon cell perfusion (red). Insert shows cell binding data (left-right channel differential signal) from two runs with and without the anti-PGSL-1 blocking mAb KPL-1.

When P-selectin-Fc was immobilized on the left channel and equivalent amounts of 19Fc were coupled on the right channel, cell adhesion was only noted on the left channel. This confirms the specificity of the measured interaction (Fig. 2).

Figure 2. Specificity - Left channel contained immobilized P-selectin-Fc and right channel contained immobilized 19-Fc. Specific binding was noted in the left channel only. Absolute signal for both channels is shown. Two cycles of cell binding were conducted with a dissociation step of between 90 and 180s. Cell binding slope was non-zero during the dissociation phase, presumably due to the spreading of cells after initial contact. Cell binding signal was ~ 2 times higher in the first binding cycle due to the availability of more unbound ligand (P-selectin).

Conclusions

- Results indicate that the measurement of cell adhesion can be easily performed using the SR7500DC unit.
- Such assays will be helpful for studies of leukocyte-endothelial cell adhesion and screening of related inhibitors.
- In general, the study demonstrates the ability to perfuse live cells and measurement of protein-carbohydrate interactions in the time scale of typical SPR runs.
- Following cell binding, cell spreading may be measured in the dissociation phase, where buffer is being perfused, in the absence of additional cells. In this way, SPR allows studies at short length and time scales that are not feasible using conventional light and fluorescence spectroscopy.

Potential Opportunities/ Future Directions

- While the current study examines binding to P-selectin versus a control substrate with no target-ligand, it is possible to immobilize another ligand like E-selectin in the second SPR channel. In this case, comparative time and spatial measurements of cell binding to two competing target-ligands can be measured simultaneously and quantitatively.
- Extension of the methods presented to other systems that involve cell adhesion like platelet adhesion under flow should be feasible.
- Methods to regenerate the sensor chip are important since this can enable reuse of the sensor chip for testing of different types of cells, combinatorial testing of drugs, and estimation of binding/unbinding rate constants.

Learn more about how Reichert pushes the limits of detection and sensitivity in label-free interaction. To improve the quality of your protein interaction research, visit www.ReichertSPR.com

®*Trademark of Reichert Technologies Company Notice: No freedom from infringement of any patent owned by Reichert or others is to be inferred. Because use conditions and applicable laws may differ from one location to another and may change with time, Customer is responsible for determining whether products and the information in this document are appropriate for Customer's use and for ensuring that Customer's workplace and disposal practices are in compliance with applicable laws and other government enactments. The product shown in this literature may not be available for sale and/or available in all geographies where Reichert is represented. The claims made may not have been approved for use in all countries. Reichert assumes no obligation or liability for the information in this document. References to "Reichert" or the "Company" mean the Reichert legal entity selling the products to Customer unless otherwise expressly noted. NO WARRANTIES ARE GIVEN; ALL IMPLIED WARRANTIES OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE ARE EXPRESSLY EXCLUDED.

Corporate Office
Reichert Technologies
3362 Walden Avenue
Buffalo, New York 14043, USA
Tel: +1 716-686-4500
Fax: +1 716-686-4555
Toll Free USA: 1-888-849-8955
reichertspr.lifesciences@ametek.com

European Service Center
Carl-von-Linde Str. 42-85716
Unterschleissheim/Munich, Germany
Tel: +49 89 315 8911 0
Fax: +49 89 315 891

www.reichertspr.com